

TITLE

Conjugation of Biomolecules Using Diels-Alder Cycloaddition

FIELD OF THE INVENTION

The invention is in the field of bioorganic chemistry, more specifically the field of conjugation of biomolecules. The conjugated products prepared by the methods of the invention are useful, for example, as inoculants for the generation of antibodies, and as vaccines. The methods of the invention may also be used to immobilize biomolecules on solid supports. The immobilized biomolecules are useful in many fields, such as for example catalysis, separation, analysis, and diagnostics.

BACKGROUND

The conjugation of biomolecules to solid and gel supports is a common operation in many laboratories, and many methods have been developed for this purpose. Immobilization of enzymes (I. Chernukhin, E. Klenova, *Anal. Biochem.* 2000, **280**:178-81), oligonucleotides (J. Andreadis; L. Chrisey, *Nucleic Acids Res.* 2000, **28**:e5; A. Drobyshev *et al.*, *Nucl. Acids. Res.* 1999, **27**:4100-4105), antibodies (P. Soltys, M. Etzel, *Biomaterials* 2000, **21**:37-48), and antigens (M. Oshima, M. Atassi, *Immunol. Invest.* 1989, **18**:841-851) on solid and gel supports enables the preparation of useful products such as chromatographic media (*Meth. Enzym.*, W. Jakoby, M. Wilchek, eds., 1974, **34**, Academic Press, NY), catalysts (T. Krogh *et al.*, *Anal. Biochem.*, 1999, **274**:153-62), biosensors (J. Spiker, K. Kang, *Biotechnol. Bioeng.* 1999, **66**:158-63), and numerous diagnostic (G. Ramsay, *Nature Biotechnol.*, 1998, **16**:40-44) and research tools (C. Bieri *et al.*, *Nature Biotechnol.* 1999, **17**:1105-1108). Even whole cells may be immobilized by such methods (E. Olivares, W. Malaisse, *Int. J. Mol. Med.* 2000, **5**:289-290).

The most robust form of attachment of a biomolecule to a surface or other support is via covalent bonds. Typically, such bonds are heteroatom-based (e.g., amide, ester, and disulfide bonds), because such bonds are easily formed under mild conditions. Non-covalent attachment via specific binding pairs (e.g., biotin-avidin or antibody-antigen interactions) is also commonly employed, but such methods still require initial conjugation of the specific binding pairs to the biomolecule and support. The use of carbon-carbon bonds for

this purpose is very rare, because formation of C-C bonds is more difficult, especially under the mild aqueous conditions appropriate for working with proteins.

The use of the Diels-Alder reaction to attach a member of a specific binding pair has been described. In this report (M.N. Yousaf and M. Mrksich, *J. Am. Chem. Soc.*, 1999, **121**:4286), a Diels-Alder reaction was used to covalently attach a biotinylated diene to an immobilized dienophile, and the immobilized biotin was subsequently used to non-covalently immobilize streptavidin. These workers have more recently used a Diels-Alder reaction to immobilize the peptide RGDS on a self-assembled alkanethiol monolayer on a gold surface (M.N. Yousaf, B.T. Houseman, M. Mrksich, *Angew. Chem. Int. Ed. Engl.*, 2001, **40**:1093). The use of the Diels-Alder reaction to effect the actual covalent coupling or immobilization event of large biomolecules, however, had not previously been described.

The conjugation of biomolecules to one another is likewise a very common procedure, and is subject to most of the concerns and limitations described above for biomolecule immobilization. Covalent attachment of haptens to proteins has been a target of synthetic endeavors since the discovery by Landsteiner that this process can convert non-immunogenic molecules to immunogenic materials (K. Landsteiner, H. Lampl, *Biochem. Zeitschr.* 1918, **86**:343). The application of this concept to carbohydrates by Goebel and Avery revealed that covalent carbohydrate-protein conjugates are immunogenic and can generate anti-carbohydrate antibodies (W. Goebel, *J. Exp. Med.* 1940, **72**:33). The use of Landsteiner's principle has led to the development of carbohydrate-protein conjugates that are valuable tools in glycomedical research, and that are useful as pharmaceuticals. In particular, protein conjugates of fragments of the capsular polysaccharide of *Haemophilus influenzae* type b have become established as successful vaccines (J. Robbins *et al.*, *J. Am. Med. Assoc.* 1996, **276**:1181). Several other bacterial saccharide-protein conjugates are in various stages of clinical studies (E. Konadu *et al.*, *J. Infect. Dis.* 1998, **177**:383-387; E. Konadu *et al.*, *Infect. Immun.* 2000, **68**:1529-1534) while numerous others are in the pre-clinical phase (V. Pozsgay *et al.*, *Proc. Natl. Acad. Sci. USA* 1999, **96**:5194).

The choice of methods for covalent bond formation between biomolecules such as carbohydrates and proteins is restricted by their limited solubility in organic solvents, and in many cases by their pH and temperature sensitivity. In almost all cases, water is the only

solvent that can be used for conjugation of carbohydrates or proteins, and the conditions are usually limited to temperatures under 50 °C and pH values between 6 and 8.

Numerous methods have been developed for the attachment of polysaccharides to proteins (C. Peeters *et al.*, in *Vaccine Protocols*, A. Robinson *et al.*, Eds., 1996 Humana Press, NJ, p. 111; W. Dick, Jr., M. Beurret, in *Contrib. Microbiol. Immunol.*, J. Cruse and R. Lewis, eds., 1989, 10:48-114, Karger, Basel; H. Jennings, R. Sood, in *Neoglycoconjugates. Preparation and Applications*, Y. Lee, R. Lee, eds., Academic Press, New York, 1994, p. 325). However, only a few of these methods are capable of coupling oligosaccharides to carriers in a site-selective fashion. Most prominent among these is reductive amination, which converts the reducing-end residue of the polysaccharide into a polyhydroxy alkylamino moiety, which unfortunately causes the loss of this unit as a true carbohydrate in the resulting glycoconjugate (V. Pozsgay, *Glycoconjugate J.* 1993, 10:133).

This problem can be solved by chemical synthesis of oligosaccharide glycosides with aglycons that bear a (latent) reactive group. Examples include alkenyl groups (M. Nashed, *Carbohydr. Res.* 1983, 123:241-246; J. Allen, S. Danishefsky, *J. Am. Chem. Soc.* 1999, 121:10875), 3-aminopropyl (G. Veeneman *et al.*, *Tetrahedron* 1989, 45:7433), 4-aminophenylethyl (R. Eby, *Carbohydr. Res.* 1979, 70:75), 4-aminophenyl (S. Stirn *et al.*, *Justus Liebigs Ann. Chem.* 1966, 696:180), 6-aminohexyl (J. Hermans *et al.*, *Rec. Trav. Chim. Pays-Bas* 1987, 106:498; R. Lee *et al.*, *Biochemistry* 1989, 28:1856), 5-methoxycarbonylpentyl (S. Sabesan, J. Paulson, *J. Am. Chem. Soc.* 1986, 108:2068; V. Pozsgay, *Org. Chem.* 1998, 63:5983), 8-methoxycarbonyloctyl (R. Lemieux *et al.*, *J. Am. Chem. Soc.* 1975, 97, 4076; B. Pinto *et al.*, *Carbohydr. Res.* 1991, 210, 199) 4-aminobenzyl (W. Goebel, *J. Exp. Med.* 1940, 72:33), ω-aldehydoalkyl (V. Pozsgay, *Glycoconjugate J.* 1993, 10:133), 3-(2-aminoethylthio)propyl (Y. Lee, R. Lee, *Carbohydr. Res.* 1974, 37:193), 2-chloroethylthioglycosides (M. Ticha *et al.*, *Glycoconjugate J.* 1996, 13:681) and 1-O-succinimide derivatives (M. Andersson, S. Oscarson, *Bioconjugate Chem.* 1993, 4:246; B. Davis, *J. Chem. Soc. Perkin I* 1999, 3215).

These aglycons introduce spacers that can be linked to a protein either directly or after insertion of a secondary linker. For this purpose the use of an activated dicarboxylic acid has been reported (R. van den Berg *et al.*, *Eur. J. Org. Chem.* 1999, 2593-2600). In

another procedure, a sulfhydryl group at the terminal position of the spacer allows the formation of a disulfide bridge with proteins using the dithiopyridyl method (J. Evenberg *et al.*, *J. Infect. Dis.* 1992, **165**(sup. 1):S152). In a related protocol, a thiolated protein is coupled with a maleimido-derivatized saccharide (J. Mahoney, R. Schnaar, *Methods Enzymol.* 1994, **242**:17). *N*-acryloylamidophenyl glycosides may be coupled to unmodified proteins using a Michael addition (A. Romanowska *et al.*, *Methods Enzymol.* 1994, **242**:90). As an alternative to glycoside formation, direct coupling of a carbohydrate to a linker via amide bonds has also been used (A. Fattom *et al.*, *Infect. Immun.* 1992, **60**:584-589), but this approach is limited to carboxyl-containing carbohydrates.

The yields of any of these methods rarely exceed 40%, and are generally in the 10-20% range (R. van den Berg *et al.*, *Eur. J. Org. Chem.* 1999, 2593-2600), especially when medium or high carbohydrate loading in the conjugate is attempted. This problem is compounded by the fact that the oligosaccharide haptens, usually obtained in multistep syntheses or by controlled degradation of polysaccharides, can rarely be recovered in their active or activable form after the coupling procedure. An additional problem with most chemical coupling methods employed to date is the formation of cross-linked byproducts, due to the presence of multiple reactive functional groups (*e.g.*, amines, acids, hydroxyls, and sulfhydryls) on most biomolecules. Avoidance of this problem requires that the reactive groups be blocked, which requires additional processing steps and may alter the physicochemical and immunological properties of the biomolecule. Thus, there remains a need for a mild and site-selective method for coupling biomolecules to one another, which avoids the problems of low yields, crosslinking, and loss of starting materials. For similar reasons there remains a need for mild and selective methods for attaching biomolecules to surfaces and solid and gel supports.

BRIEF DESCRIPTION OF THE INVENTION

The present invention provides an experimentally simple protocol for the covalent attachment of biomolecules to one another and to supports, that can avoid many of the above-mentioned problems. The invention makes use of the well-known Diels-Alder cycloaddition reaction that takes place between a double bond and a conjugated diene. This reaction has traditionally been carried out in organic solvents, but can proceed in aqueous

solutions as well (R. Breslow, D. Ridcut, *J. Am. Chem. Soc.* 1980, **102**:7816; A. Lubineau, J. Auge, *Top. Curr. Chem.* 1999, **206**:1; P. Garner, in *Organic Synthesis in Water*, P. Grieco, ed., Blackie Academic and Professional, London, 1998, p. 1.)

Carbohydrates have been employed as chiral auxiliaries and/or water solubilizing agents for Diels-Alder reactions, wherein a conjugated diene system is converted to a glycoside prior to the cycloaddition (A. Lubineau *et al.*, *J. Chem. Soc. Perkin I* 1997, 2863-2867; see also S. Pellegrinet, R. Spanevello, *Org. Lett.* 2000, **2**:1073-1076). As noted above, the Diels-Alder reaction has also been used to covalently attach biotin to a support (M.N. Yousaf and M. Mrksich, *J. Am. Chem. Soc.*, 1999, **121**:4286). However, the Diels-Alder reaction has not previously been extended to the direct covalent conjugation of biopolymers or other types of polymeric materials. Among the advantages of the method of the invention are the mild and neutral conditions, good yields, negligible cross-linking, and facile recovery of excess and/or unreacted biomolecules in their conjugatable form.

The invention also provides conjugated biomolecules, which are useful as immunostimulatory agents for production of antibodies and induction of immunity, methods of inducing antibody production with the conjugated biomolecules, and vaccine compositions comprising the conjugated biomolecules.

The invention also provides polyclonal and monoclonal antibodies generated by administration of the conjugated biomolecules to a mammal, and methods of using the induced antibodies for inducing passive immunity. The antibodies are useful of therapeutic, diagnostic, and analytical purposes.

The invention also provides immobilized biomolecules, and methods for their preparation, which are useful in many areas, such as chromatographic media, catalysts, components of diagnostic devices, biosensors, and as research tools.

DETAILED DESCRIPTION OF THE INVENTION

The invention provides a new method for conjugation of biomolecules based on the Diels-Alder cycloaddition reaction. The technique involves the introduction of an activated double bond into a first biomolecule component, and a conjugated diene into a second biomolecule component, which are to be covalently linked together. The diene- and dienophile-modified biomolecules may then be purified to the extent desired. The two

components are then simply combined under neutral conditions, and the cycloaddition reaction is allowed to proceed.

As used herein, the term "biomolecule" refers generally to the large, complex molecules produced within living cells, and to synthetic and semi-synthetic analogues thereof. Examples include proteins, peptides, oligo- and poly-saccharides, and oligo- and poly-nucleic acids, and various combinations thereof such as for example glycoproteins and nucleoproteins. Larger complexes, such as ribosomes, cellular substructures, and even entire cells are intended to fall within the meaning of the term as well.

In certain of the examples presented, the diene moiety is introduced into a carbohydrate component and the activated double bond into a polypeptide component. This may be reversed if desired, as a matter of convenience or if required by the synthetic design, as shown in another of the examples. The cycloaddition step proceeds in most cases under neutral conditions, at or below physiological temperatures. Where they are of sufficiently low molecular weight, unreacted components can be recovered for re-use by simple diafiltration of the coupling reaction mixture.

The embodiment of the invention disclosed in certain of the examples below incorporates an electron-deficient carbon-carbon double bond into a protein, with human serum albumin (HSA) being used as an example and the commercially available reagent 3-sulfosuccinimidyl 4-maleimidobutyrate being used as the reagent. In these embodiments, the diene component of the Diels-Alder reaction is incorporated into a carbohydrate, with derivatives of *trans,trans*-hexa-2,4-dien-1-ol, 1-amino-hexa-2,4-diene, and octa-2,4-dienoic acid hydrazide being used as dienes. It will be understood that in general, the Diels-Alder reaction will occur between sterically accessible dienes and dienophiles regardless of the nature of the attached biomolecules, and that by appropriate selection of reagents both homoconjugates and heteroconjugates of proteins, carbohydrates, and oligonucleotides can be carried out by the methods of this invention.

The methods of the invention unexpectedly provide for a degree of control over the rate of coupling of the diene and dienophile components, simply by modification of the linker moieties. The molecular weight of the conjugate approaches a limiting value as the coupling reaction proceeds, which depends on the mass of the carbohydrate being attached.

The observed increase in molecular weight as a function of time may therefore be fit to a pseudo-first order reaction kinetic equation of the form

$$\Delta mw = \Delta mw_{\max}(1 - e^{-kt})$$

where Δmw is the increase in molecular weight, Δmw_{\max} is the maximum increase that could be obtained if all the protein-linked starting material were to react, k is the rate constant, and t is time. The curves drawn through the data of Figures 1 and 2 (see Example 5 for the experimental details) were obtained by fitting the parameters of this equation to the data. The results of this fitting are summarized in Table 2.

The faster incorporation of construct **27** relative to **8** is an interesting observation. The observed difference apparently correlates with the distance between the hydrophilic rhamnose moiety and the hydrophobic diene part of the molecule. According to Breslow, the accelerating effect of water on the rate of Diels-Alder reactions is due to hydrophobic packing of the reactants (R. Breslow, D. Rideout, *J. Am. Chem. Soc.* 1980, **102**:7816). Such an effect should be more pronounced in compound **27**, where the diene sector is more isolated from the hydrophilic portion, than in compound **8**.

The Diels-Alder reaction requires a highly organized transition state, and the ability of the components of the present invention to achieve the proper geometry for cycloaddition despite the mass and bulk of the attached biomolecules is remarkable.

In a preferred embodiment of the invention, one of the biomolecules to be linked is a hapten or antigen, and the other is a carrier. In a particularly preferred embodiment, the hapten or antigen is a polysaccharide moiety. Examples of antigenic polysaccharides are the capsular polysaccharides of *Haemophilus influenzae* type b, *Neisseria meningitidis*, Group B *Streptococci*, *Salmonella typhi*, *E. coli*, and *Pneumococci*.

Carriers are chosen to increase the immunogenicity of the hapten or antigen, and/or to raise antibodies against the carrier itself which may be medically beneficial. Carriers that fulfill these criteria are known in the art (see, e.g., A. Fattom *et al.*, *Infect. Immun.* 1990, **58**, 2309-2312; Devi, J. Robbins, R. Schneerson, *Proc. Natl. Acad. Sci. USA* 1991, **88**:7175-7179; S. Szu, X *et al.*, *Infect. Immun.*, 1991, **59**:4555-4561; S. Szu *et al.*, *J. Exp. Med.*, 1987, **166**:1510-1524). A carrier can be a natural, semi-synthetic, or synthetic

material containing one or more functional groups, for example primary and/or secondary amino groups, azido groups, hydroxyl groups, or carboxyl groups, to which a diene or dienophile Diels-Alder reactant moiety can be attached. The carrier can be water soluble or insoluble, and is preferably a polypeptide.

5 Examples of water soluble polypeptide carriers include, but are not limited to, natural, synthetic, or semisynthetic peptides or proteins from bacteria or viruses, *e.g.*, bacterial, bacterial outer membrane proteins, bacterial toxins and toxoids such as tetanus toxin/toxoid, diphtheria toxin/toxoid, *Pseudomonas aeruginosa* exotoxin/toxoid/protein, pertussis toxin/toxoid, and *Clostridium perfringens* exotoxins/toxoid. Viral proteins such as
10 hepatitis B surface antigen and core antigen may also be used as carriers, as well as proteins from higher organisms such as keyhole limpet hemocyanin, horseshoe crab hemocyanin, edestin, mammalian serum albumins, mammalian gamma-globulins, and IgG.

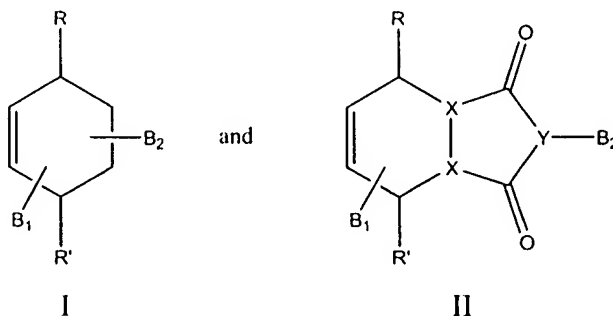
 Polysaccharide carriers include, but are not limited to, dextran, capsular polysaccharides from microorganisms such as the Vi capsular polysaccharide from *S. typhi*,
15 which is described in U.S. patent 5,204,098, (incorporated by reference herein); *Pneumococcus* group 12 (12F and 12A) polysaccharides; *Haemophilus influenzae* type d polysaccharide; and certain plant, fruit, or synthetic oligo- or polysaccharides which are immunologically similar to capsular polysaccharides, such as pectin, D-galacturonan, oligogalacturonate, or polygalacturonate, for example as described in U.S. patent 5,738,855
20 (incorporated by reference herein).

 Examples of water insoluble carriers include, but are not limited to, aminoalkyl agarose, *e.g.*, aminopropyl or aminoethyl SEPHAROSE (Pharmacia Inc., Piscataway, NJ), aminopropyl glass, cross-linked dextran, and the like, to which a diene or dienophile can be attached. Other carriers may be used provided that a functional group is available for
25 covalently attaching a diene or dienophile.

 Examples of dienophiles include, but are not limited to, maleimides, acrylamides, azodicarboxylates, quinones, and 1,2,4-triazoline-3,5-diones. Examples of dienes include, but are not limited to, esters and glycosides of hexa-2,4-dien-1-ol, penta-2,4-dien-1-ol, furan-2-methanol, and furan-1-methanol; esters, amides, and hydrazides of octa-
30 2,4-dienoic acid; and amides of 1-aminohexa-2,4-diene and 1- and 2-aminomethylfuran. The

above-mentioned amines may also be coupled with aldehydo-biomolecules via reductive amination, and hydrazides may be attached to such biomolecules via condensation.

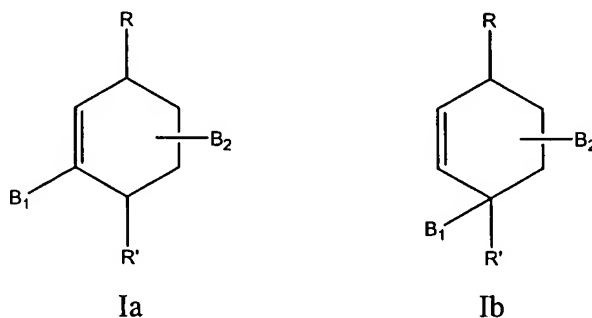
The invention also provides biomolecule conjugates of general formulas I and II below:



where R and R' are independently H or methyl, or together constitute CH₂, CH₂CH₂, SO₂, or O; X is CH or N; Y is N, CH=C, or NH-N; and B₁ and B₂ comprise biomolecules independently selected from the group consisting of polypeptides, carbohydrates, polysaccharides, and nucleic acids, and are optionally attached via a linker.

The invention also provides immobilized biomolecules of formulas I and II above, wherein one of B₁ and B₂ may be a solid or gel support. Examples of solid supports include, but are not limited to, aminopropylsilylated glass and silica surfaces, gold surfaces functionalized with thiol-bound linkers, functionalized macroporous polystyrene beads, and surface-derivatized microtiter plate wells. Examples of gel supports include, but are not limited to, functionalized agarose gels such as cyanogen bromide activated agarose, aminoethyl agarose, and carboxymethyl agarose.

The formulas above are intended to indicate that the group B₁ may be attached alpha or beta to the group R' as shown below:



There are many known methods of attachment of small molecules to biomolecules, there are many known linker moieties for attachment of chemical moieties to biomolecules, and there are many known dienes and dienophiles that readily take part in cycloaddition reactions at or near room temperature. Those skilled in the art will thus appreciate that there are many obvious combinations of attachment methods, linkers, and diene and dienophile partners that may be employed in the method of biomolecule coupling disclosed herein, which are equivalent to the examples provided. Such modifications of the disclosed methods and resulting compositions are intended to be within the scope and spirit of the present invention.

It is another object of the invention to provide methods of using the polysaccharide-carrier conjugates of this invention for eliciting an immunogenic response in mammals, including but not limited to responses which provide protection against, or reduce the severity of, bacterial and viral infections. The pharmaceutical compositions of this invention are expected to be capable, upon injection into a mammal, of inducing serum antibodies against the polysaccharide component of the conjugate.

The invention also provides methods of using such conjugates, and/or pharmaceutical compositions comprising such conjugates, to induce in mammals, in particular, humans, the production of antibodies which immunoreact with the polysaccharide component of the conjugates. Antibodies which immunoreact with a bacterial or viral polysaccharide are useful for the identification or detection of microorganisms expressing the polysaccharide, and/or for diagnosis of infection. Antibodies against the polysaccharide may be useful in increasing resistance to, preventing, ameliorating, and/or treating illnesses caused by microorganisms or viruses that express the polysaccharide.

The compositions of this invention are intended for active immunization for prevention of infection, and for preparation of immune antibodies. The compositions of this invention are designed to induce antibodies specific to microorganisms expressing the polysaccharide component of the conjugate, and to confer specific immunity against infection with such microorganisms.

This invention also provides compositions, including but not limited to, mammalian serum, plasma, and immunoglobulin fractions, which contain antibodies which

are immunoreactive with the polysaccharide component of the conjugates of this invention, and which preferably also contain antibodies which are immunoreactive with the protein component. These antibodies and antibody compositions may be useful to prevent, treat, or ameliorate infection and disease caused by the microorganism. The invention also provides
5 such antibodies in isolated form. The invention further provides methods of inducing in mammals antibodies which immunoreact with a polysaccharide, the methods comprising administering to a mammal a composition of the invention.

The invention also provides monoclonal antibodies, preferably produced by hybridomas, which immunoreact with a polysaccharide. The nucleic acid sequences encoding
10 these antibodies are obtained from a mammal in which the production of anti-polysaccharide antibodies has been induced by administering a composition of the invention.

As used herein, the terms "immunoreact" and "immunoreactivity" refer to specific binding between an antigen or antigenic determinant-containing molecule and a molecule having an antibody combining site, such as a whole antibody molecule or a portion
15 thereof.

As used herein, the term "antibody" refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules. Exemplary antibody molecules are intact immunoglobulin molecules, substantially intact immunoglobulin molecules and portions of an immunoglobulin molecule, including those portions known in
20 the art as Fab, Fab', F(ab')₂ and F(v), as well as chimeric antibody molecules.

Polymeric carriers

Carriers are chosen to increase the immunogenicity of the polysaccharide and/or to raise antibodies against the carrier which are medically beneficial. Carriers that fulfill these criteria are well-known in the art. A polymeric carrier can be a natural or a
25 synthetic material containing one or more functional groups, for example primary and/or secondary amino groups, azido groups, or carboxyl groups, to which a diene or dienophile component can be attached. Carriers can be water soluble or insoluble. The examples below employ proteins as carriers.

Regardless of the precise method used to prepare the conjugate, after the Diels-
30 Alder coupling reaction has been carried out the unreacted materials are preferably removed

by routine physicochemical methods, such as for example dialysis, gel filtration or ion exchange column chromatography, depending on the materials to be separated. The final conjugate consists of the polysaccharide and the carrier bound through a Diels-Alder adduct.

Dosage for Vaccination

5 The present inoculum contains an effective, immunogenic amount of a polysaccharide-carrier conjugate. The effective amount of polysaccharide-carrier conjugate per unit dose sufficient to induce an immune response depends, among other things, on the immunogenicity of the polysaccharide, the species of mammal inoculated, the body weight of the mammal, and the chosen inoculation regimen, as is well known in the art. Inocula
10 typically contain polysaccharide-carrier conjugates with concentrations of polysaccharide from about 1 micrograms to about 500 micrograms per inoculation (dose), preferably about 3 micrograms to about 50 micrograms per dose, and most preferably about 5 micrograms to 25 micrograms per dose.

 The term "unit dose" as it pertains to the inocula refers to physically discrete
15 units suitable as unitary dosages for mammals, each unit containing a predetermined quantity of active material (polysaccharide) calculated to produce the desired immunogenic effect in association with the required diluent.

 Inocula are typically prepared in physiologically and/or pharmaceutically tolerable (acceptable) carriers, and are preferably prepared as solutions in physiologically
20 and/or pharmaceutically acceptable diluents such as water, saline, phosphate-buffered saline, or the like, to form an aqueous pharmaceutical composition. Adjuvants, such as aluminum hydroxide, QS-21, TiterMax™ (CytRx Corp., Norcross GA), Freund's complete adjuvant, Freund's incomplete adjuvant, interleukin-2, thymosin, and the like, may also be included in the compositions.

25 The route of inoculation may be by intramuscular or subcutaneous injection or the like, so long as it results in eliciting antibodies reactive against the polysaccharide component. It is anticipated that in some cases the composition can be administered orally or intranasally, for example when mucosal immunity is to be induced. In order to increase the antibody level, a second or booster dose may be administered approximately 4 to 6 weeks
30 after the initial administration. Subsequent doses may be administered as deemed necessary

by the practitioner.

Antibodies

An antibody of the present invention is typically produced by immunizing a mammal with an immunogen or vaccine containing a polysaccharide-carrier conjugate, preferably a polysaccharide-protein conjugate, to induce in the mammal antibody molecules having immunospecificity for the polysaccharide component of the conjugate. Antibody molecules having immunospecificity for the protein carrier may also be produced. The antibody molecules may be collected from the mammal and, optionally, isolated and purified by methods known in the art.

Human or humanized monoclonal antibodies are preferred, including but not limited to those identified by phage display technology, and including but not limited to those made by hybridomas and by mice with human immune systems or human immunoglobulin genes. The antibody molecules of the present invention may be polyclonal or monoclonal. Monoclonal antibodies may be produced by methods well-known in the art. Portions of immunoglobulin molecules, such as Fabs, may also be produced by methods known in the art.

An antibody of the present invention may be contained in blood plasma, serum, hybridoma supernatants and the like. Alternatively, the antibodies of the present invention are isolated to the extent desired by well known techniques such as, for example, ion chromatography or affinity chromatography. The antibodies may be purified so as to obtain specific classes or subclasses of antibody such as IgM, IgG, IgA, IgG₁, IgG₂, IgG₃, IgG₄ and the like. Antibodies of the IgG class are preferred for conferring passive immunity. The antibodies of the present invention have a number of diagnostic and therapeutic uses. The antibodies can be used as an *in vitro* diagnostic agents to test for the presence of microorganisms in biological samples or in water or food samples, in standard immunoassay protocols. Such assays include, but are not limited to, agglutination assays, radioimmunoassays, enzyme-linked immunosorbent assays, fluorescence assays, Western blots and the like. In one such assay, for example, the sample is contacted with first antibodies of the present invention, and a labeled second antibody is used to detect the presence of polysaccharides to which the first antibodies have bound.

Such assays may be, for example, of direct format (where the labeled first

antibody is reactive with the polysaccharide), an indirect format (where a labeled second antibody is reactive with the first antibody), a competitive format (such as the addition of a labeled polysaccharide), or a sandwich format (where both labeled and unlabelled antibody are utilized), as well as other formats described in the art.

5 In providing the antibodies of the present invention to a recipient mammal, the dosage of administered antibodies will vary depending upon such factors as the mammal's age, weight, height, sex, general and specific medical conditions, and the like.

In general, it is desirable to provide the recipient with a dosage of antibodies which is in the range of from about 1 mg/kg to about 10 mg/kg body weight of the mammal, 10 although a lower or higher dose may be administered. The antibodies of the present invention are intended to be provided to the recipient subject in an amount sufficient to prevent, or lessen or attenuate the severity, extent or duration of the infection.

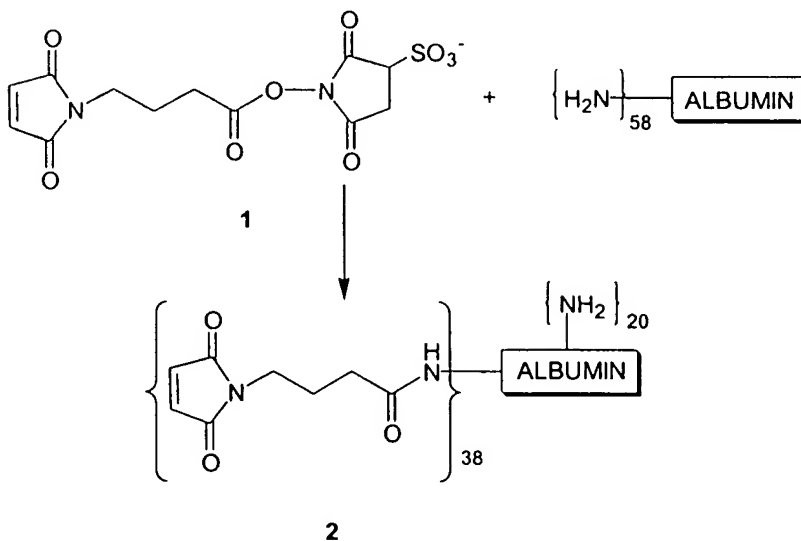
In order to facilitate the administration of the conjugates of the invention to mammals, it is preferred that the conjugate be formulated with a pharmaceutically acceptable 15 carrier. (Those skilled in the art will appreciate that the term "carrier," when used in this context, has a different meaning than when it is used to refer to a biomolecule component of the conjugate.) Examples of pharmaceutically acceptable carriers include sterile water and saline, both of which may be buffered with phosphate, citrate, and the like. The conjugates of the invention may be provided in solution or suspension in a pharmaceutically acceptable 20 carrier, or they may be provided in dry form and reconstituted with the pharmaceutically acceptable carrier prior to administration.

The administration of the conjugates and compositions of the invention may be for prophylactic or therapeutic purposes. When provided prophylactically, the agents are provided in advance of any symptom. The prophylactic administration of the agent serves to 25 prevent or ameliorate any subsequent infection. When provided therapeutically, the agent is provided at (or shortly after) the onset of a symptom of infection.

For all therapeutic, prophylactic and diagnostic uses, the polysaccharide-carrier conjugates of this invention, as well as antibodies and other necessary reagents and appropriate devices and accessories may be provided in kit form so as to be readily available 30 and easily used.

The examples below will be understood to be merely representative of the invention, and are not intended to limit the scope of the appended claims in any way.

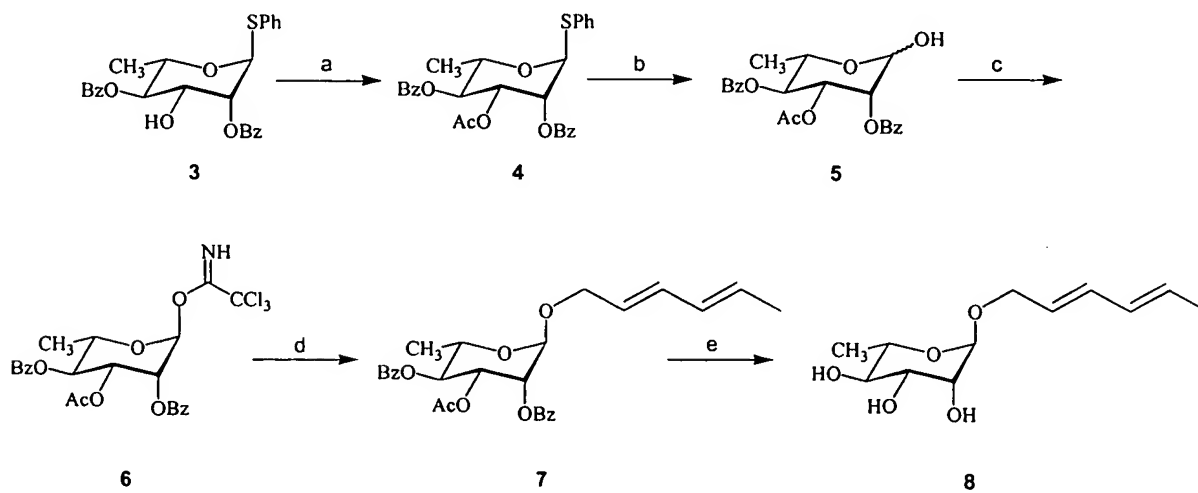
EXAMPLE 1



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Dieneophile component 2: Treatment of human serum albumin (HSA) with a 1.6 molar excess (based on 58 available amino groups) of 3-sulfosuccinimidyl 4-maleimidobutyrates (1) in a pH 7.5 phosphate buffer afforded the intermediate 2, which contained an average of 38 maleimido moieties per protein molecule, as indicated in the

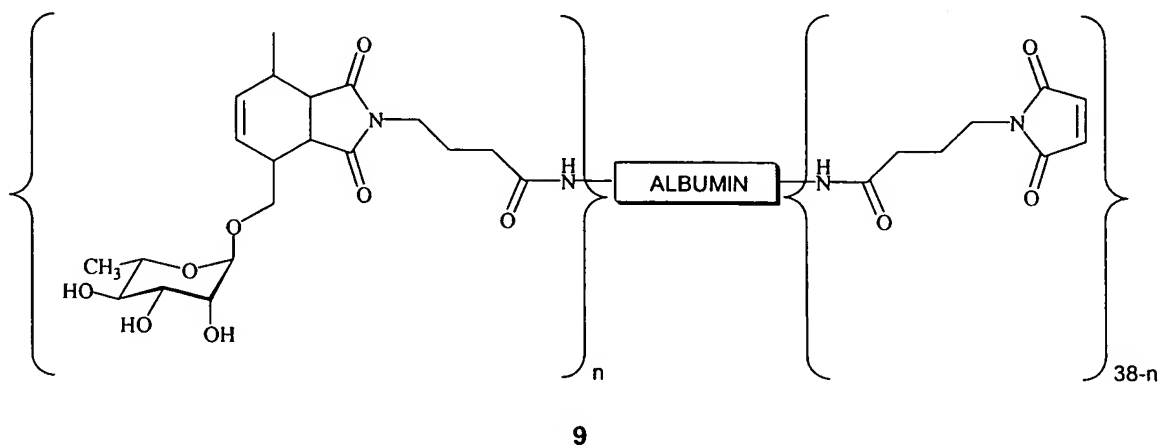
10 formula (determined by MALDI-TOF mass spectroscopy).



Diene component 8: The phenylthio rhamnoside **3** was prepared as described previously (V. Pozsgay, *Carbohydr. Res.*, 1992, **235**:295). Rhamnoside **3** was treated with acetic anhydride and pyridine to afford **4**, ^1H NMR (CDCl_3 , δ) 8.11-7.26 (m, 15 H), 5.79 (dd, 1 H), 5.64-5.52 (m, 3 H), 5.53 (t, 1 H, $J = 10.0$ Hz), 4.69 (dq, 1 H), 1.89 (s, 2 H), 1.35 (d, 3 H, $J = 6.3$ Hz), ^{13}C (CDCl_3 , δ) 170.1, 165.7, 165.5, 133.5-127.9, 85.8, 72.1, 71.9, 69.5, 68.1, 20.6, 17.6.

From **4** the hemiacetal **5** was obtained by hydrolysis with mercuric trifluoroacetate (L. Yan, D. Kahne, *J. Am. Chem. Soc.* 1996, **118**:9239), ^1H NMR (CDCl_3 , δ) 8.1-7.4 (m, 10 H), 5.71 (dd, 1 H, $J = 3.4$ Hz, $J = 9.9$ Hz), 5.78 (dd, 1 H), 5.49 (t, 1 H, $J = 9.9$ Hz), 5.38 (br d, 1 H), ^{13}C (CDCl_3 , δ) 170.3, 165.8, 92.2, 71.9, 71.0, 68.9, 66.7, 20.7, 17.7.

Hemiacetal **5** was converted to the trichloroacetimidate **6**, and glycosylation of *trans,trans*-hexa-2,4-dien-1-ol with **6** using $\text{CF}_3\text{SO}_3\text{Si}(\text{CH}_3)_3$ as the activator afforded the glycoside **7**. The acetyl groups were then removed by treatment with NaOMe to afford the diene rhamnoside **8**. ^1H NMR (D_2O , δ) 6.33 (dd, 1 H, $J = 9.5$ Hz, $J = 10.2$ Hz), 6.16 (ddd, 1 H, $J = 1.6$ Hz, $J = 10.2$ Hz, $J = 14.8$ Hz), 5.85 (m, 1 H), 5.69 (m, 1 H), 4.83 (d, 1 H, $J = 1.7$ Hz), 4.21 (dd, 1 H, $J = 6.4$ Hz, $J = 12.4$ Hz), 4.07 (dd, 1 H, $J = 7.1$ Hz, $J = 12.4$ Hz), 3.91 (dd, 1 H, $J = 1.7$ Hz, $J = 3.4$ Hz), 3.69 (dq, 1 H), 3.73 (dd, 1 H, $J = 3.4$ Hz, $J = 9.6$ Hz), 3.44 (t, 1 H, $J = 9.6$ Hz), 1.75 (d, 3 H, $J = 6.8$ Hz), 1.28 (d, 3 H, $J = 6.3$ Hz), ^{13}C (D_2O , δ) 136.7, 132.7, 131.1, 125.6, 99.8, 72.8, 71.1, 71.0, 70.9, 69.4, 68.6, 18.2, 17.4.



Coupling reaction: An excess of **8** was treated in an aqueous solution with the maleimido-derivatized protein **2**. The average incorporations of the hapten, as a function of time and temperature, are shown in Table 1. This data was obtained from the average molecular mass of the conjugates determined by the MALDI-TOF method. As expected for a concerted cycloaddition reaction, the incorporation level depends on the reaction time and temperature. At room temperature, approximately 63% of the available dienophile moieties in the protein participated in adduct formation within 36 h, while at 40°C almost complete utilization of these moieties occurred after four days (Table 1). The unreacted diene **8** was recovered by diafiltration, and the conjugate **9** (free amino groups not shown) was then obtained as a white solid after freeze-drying.

Table 1.

Time and temperature dependence of the cycloaddition between **2** and **8**

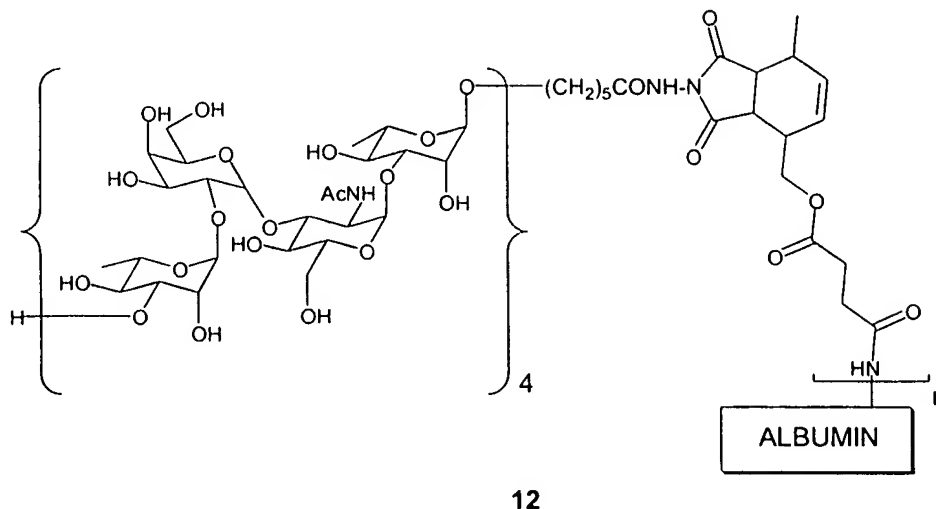
Time (h)	Composition of the conjugate (mol hapten/mol albumin)	
	22°C	40°C
36	24	27
100	28	37

EXAMPLE 2

Dienophile component: The glycoside of 6-hydroxyhexanoic acid hydrazide with the tetramer of (α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(α -D-galactopyranosyl)-(1 \rightarrow 3)-(α -D-glucopyranosyl)-(1 \rightarrow 3)- α -L-rhamnopyranose is prepared, according to the procedure disclosed in international patent application WO 99/03871. Treatment with maleic anhydride provides an N-terminal maleimide derivative **10**.

Diene component: *Trans,trans*-hexa-2,4-dien-1-ol and succinic anhydride are reacted in the presence of N,N-dimethylaminopyridine to provide *trans,trans*-hexa-2,4-dien-1-ol monosuccinate. An aqueous solution of an excess of the monosuccinate is activated with 1-

[3-(dimethylamino)propyl]-3-ethylcarbodiimide and coupled with human serum albumin, to provide a poly(diene) derivative 11.



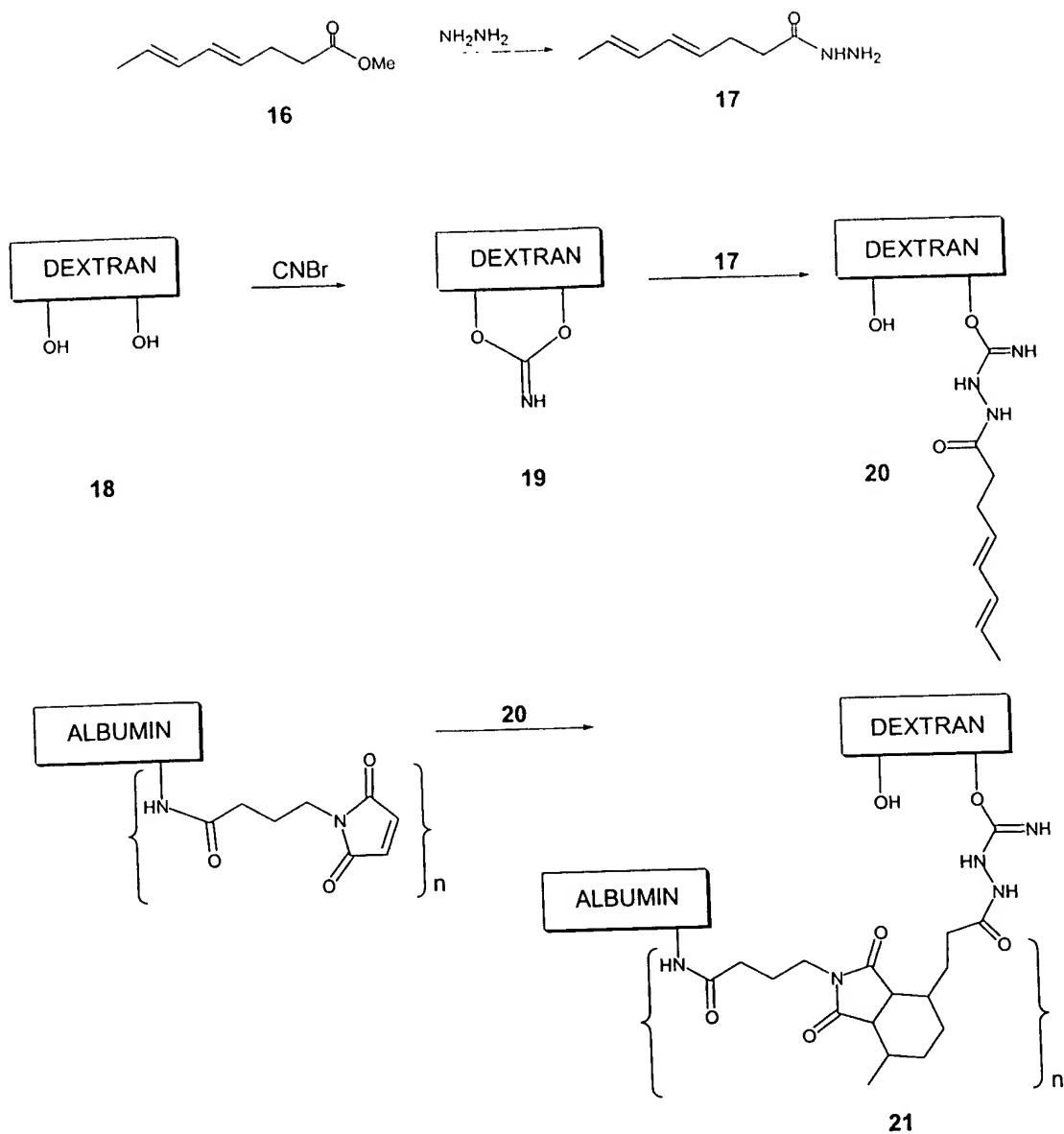
Coupling reaction: An aqueous solution of 11 and excess 10 is incubated at 35°C for 4 days, and the resulting conjugate 12 ($n \leq 58$) is purified by diafiltration and lyophilized. The conjugate 12 is expected to be useful for inducing antibodies against *Shigella dysenteriae*.

EXAMPLE 3

Diene component: The Vi capsular polysaccharide of *Salmonella typhi* (Pasteur Merieux Serums et Vaccins, Lyon FR) is dissolved in water, excess 2,4-hexadienylamine is added, and the solution buffered to pH 5.0. A slight excess of 1-ethyl-3-[3-(dimethylamino)propyl]carbodiimide in water is added. After 4 h at 37°, the mixture is adjusted to pH 7.5 and dialyzed to remove excess reagents. The resulting solution of 13 is used immediately.

Dienophile component: *Pseudomonas aeruginosa* recombinant exoprotein A (Fattom *et al.*, *Infect. Immun.* 1992, 60:584-589) is treated with excess 3-sulfosuccinimidyl 4-maleimidobutyrate (1) as in Example 1, followed by dialysis and lyophilization to provide a maleimide derivatized protein 14.

EXAMPLE 4



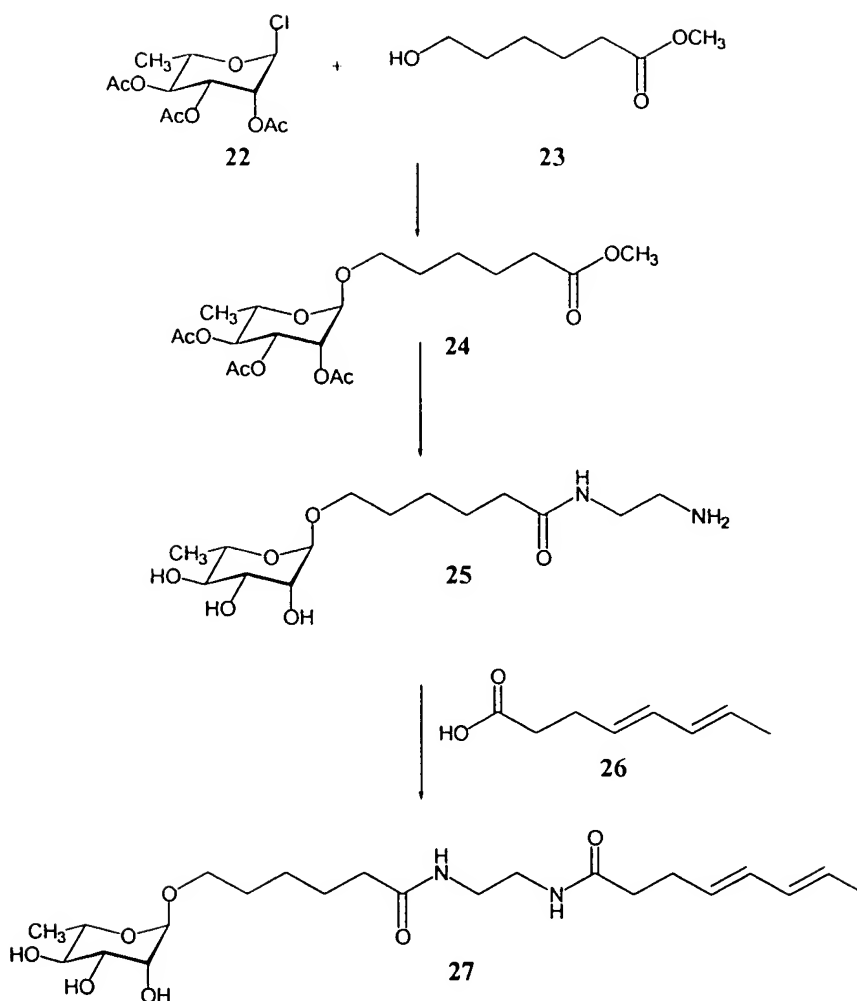
- Diene component:** To a solution of methyl octa-4,6-dienoate (**16**) (6 g) in methanol (10 ml) was added hydrazine (3 ml) at room temperature. After 24 h, the solution was diluted with water (50 ml). The crystalline precipitate was isolated by filtration to afford octa-4,6-dienoic acid hydrazide (**17**) as colorless microcrystals, yield 5.7 g. Dextran (nominal MW 10 kDa, Pharmacia) was diafiltered through a YM10 membrane (MW cutoff 10 kDa,

Millipore) using 3 changes of water. The solution that passed through the membrane was diafiltered through a YM3 (MW cutoff 3000 Da) membrane, using five changes of water. The material retained by the membrane was lyophilized. A stirred solution of the dextran **18** thus obtained (22.5 mg, 2.25 μmol , corresponding to 139 μmol of glucose) in H_2O (2.2 ml) at 5 °C (ice bath) was equipped with a temperature-sensing pH electrode, and 0.1 M NaOH was added with a 100 μL microsyringe until the pH reached 10.5. To this solution was added BrCN (50 μL of a 5 M solution in acetonitrile). The pH of the solution was maintained between 10.5 and 10.8 by addition of 0.1 M NaOH with a microsyringe. After 6 min, the solution was adjusted to pH 8.5 by addition of pH 8.0 phosphate buffer (*ca.* 1 ml). To the reaction mixture was immediately added a solution of octa-4,6-dienoic acid hydrazide (3.5 mg, 25 μmol) in 0.5 ml dimethyl sulfoxide. The pH of the reaction mixture rose to 8.95, at a temperature of 9 °C. The ice bath was removed and the stirred solution was allowed to reach 25 °C. After 2.5 h, the pH was 8.18 and remained unchanged over 5 min. The solution of **20** was diafiltered through a YM-3 membrane using 5 changes of water (4 ml each).

Dienophile component: A stirred solution of human serum albumin (11.4 mg, 0.17 μmol , 9.88 μmol of amino groups) in pH 7.5 buffer (1 ml) was treated at 5 °C (ice-bath) with 3-sulfosuccinimidyl 4-maleimidobutyrate (1) (0.52 mg, 1.36 μmol). The solution was stirred for 15 min at 5 °C then for another 15 min at room temperature followed by diafiltration through a YM-10 membrane using 5 changes of water (4 ml each).

Coupling reaction: The residual solutions containing the modified dextran **20** and the modified human serum albumin were combined. The total volume of the combined solution was approx. 1 ml. After 22 h at room temperature, the reaction mixture was diafiltered through a YM-10 membrane using 6 changes of H_2O (5 ml each), and the residue was freeze-dried. MALDI mass spectroscopy showed that most of the albumin is consumed. The conjugate **21** that is formed has average molecular weight of 90 kDa.

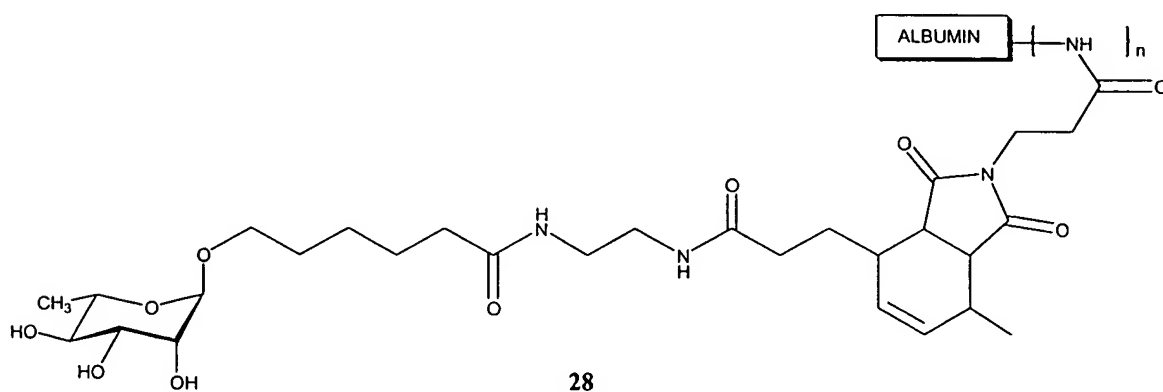
EXAMPLE 5



5 Diene component: 6-Hydroxyhexanoic acid **23** (S. Sabesan, J.C. Paulson, *J. Am. Chem. Soc.*, 1986, **108**:2068) and glycosyl chloride **22** (V. Pozsgay, *Glycoconj. J.*, 1993, **10**:133), in methylene chloride at -40 °C, were treated with silver triflate and 2,6-di-*t*-butyl-4-methylpyridine for 10 min. to afford glycoside **24**. Deprotection with a catalytic amount of NaOMe in MeOH at 23 °C for 24 hr, followed by treatment with ethylenediamine in ethanol at
 10 80 °C for 12 hr, provided **25** in 74% overall yield. ¹H NMR (500 MHz, CD₃OD): δ = 4.64 (br s, 1 H), 3.78 (dd, 1 H), 3.66 (m, 1 H), 3.55 (dq, 1 H), 3.39 (m, 1 H), 3.35 (t, 1 H), 3.36 (t, 2 H), 2.76 (t, 2 H), 2.21 (t, 2 H), 1.54-1.68 (m, 4 H), 1.35-1.46 (m, 2 H), 1.24 (d, 3 H).

Octadienoic acid **26** was prepared from the corresponding methyl ester (T. Hudlicky et al., *J. Org. Chem.*, 1980, **45**:5020). Acylation of **25** with **26** (dicyclohexylcarbodiimide, EtOAc, MeOH) afforded the glycoside diene **27** in 92% yield, ¹H NMR (500 MHz, CD₃OD): δ = 6.09 (m, 2 H), 5.70 (m, 1 H), 5.57 (m, 1 H), 4.77 (br s, 1 H), 3.92 (dd, 1 H), 3.63-3.75 (m, 4 H), 3.53 (m, 1 H), 3.43 (t, 1 H), 3.32 (m, 4 H), 2.32 (m, 4 H), 2.22 (m, 2 H), 1.72 (d, 3 H), 1.53-1.67 (m, 4 H), 1.30-1.42 (m, 2 H), 1.29 (d, 3 H).

Dienophile component: A stirred solution of human serum albumin in pH 7.5 buffer (1 ml) was treated at 5 °C (ice-bath) with 3-sulfosuccinimidyl 4-maleimidobutyrate (**1**) as described above, to provide maleimido-derivatized protein containing an average of 29 maleimido groups per molecule of protein.



Coupling reaction: The diene **27** was treated with the maleimido-derivatized protein in water at 23 °C. Samples taken at various times were diafiltered through a 10 kDa cutoff membrane then were lyophilized and subjected to MALDI-TOF mass spectrometry. The increase in molecular weight of the resulting conjugate **28** ($n \leq 29$), relative to the mass of the core protein with the dienophiles attached, is shown as a function of time in Figure 1.

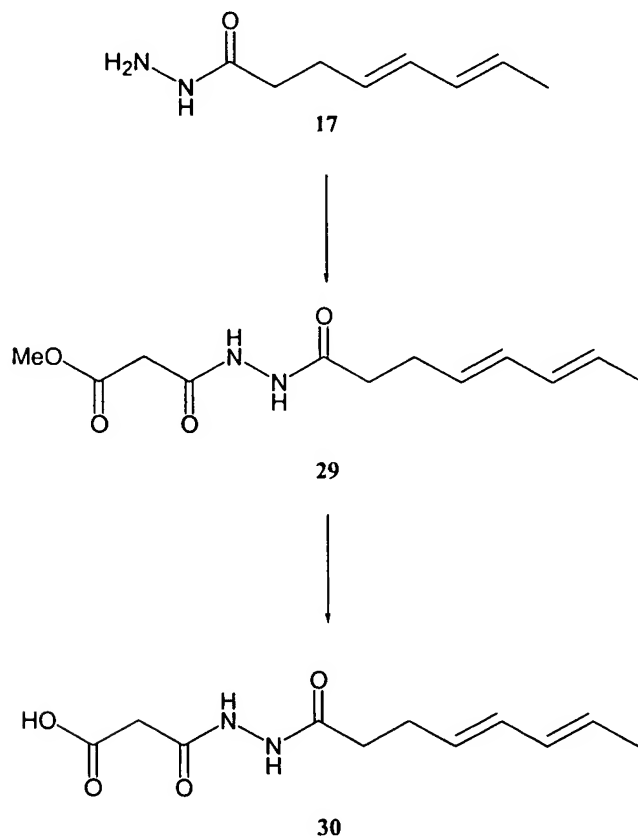
A similar time-dependent experiment was conducted with the same maleimido-derivatized protein, using previously described diene **8**, and the results are shown in Figure 2. Kinetic parameters are shown in Table 2.

Table 2
Kinetic Parameters for Conjugation Reactions

Diene	Parameters for $\Delta mw = \Delta mw_{max}(1 - e^{-kt})$		
	Δmw_{max}	$k \text{ (min}^{-1}\text{)}$	$t_{1/2} \text{ (min)}$
14	9000	0.007	99
8	4350	0.0045	154

5

EXAMPLE 6

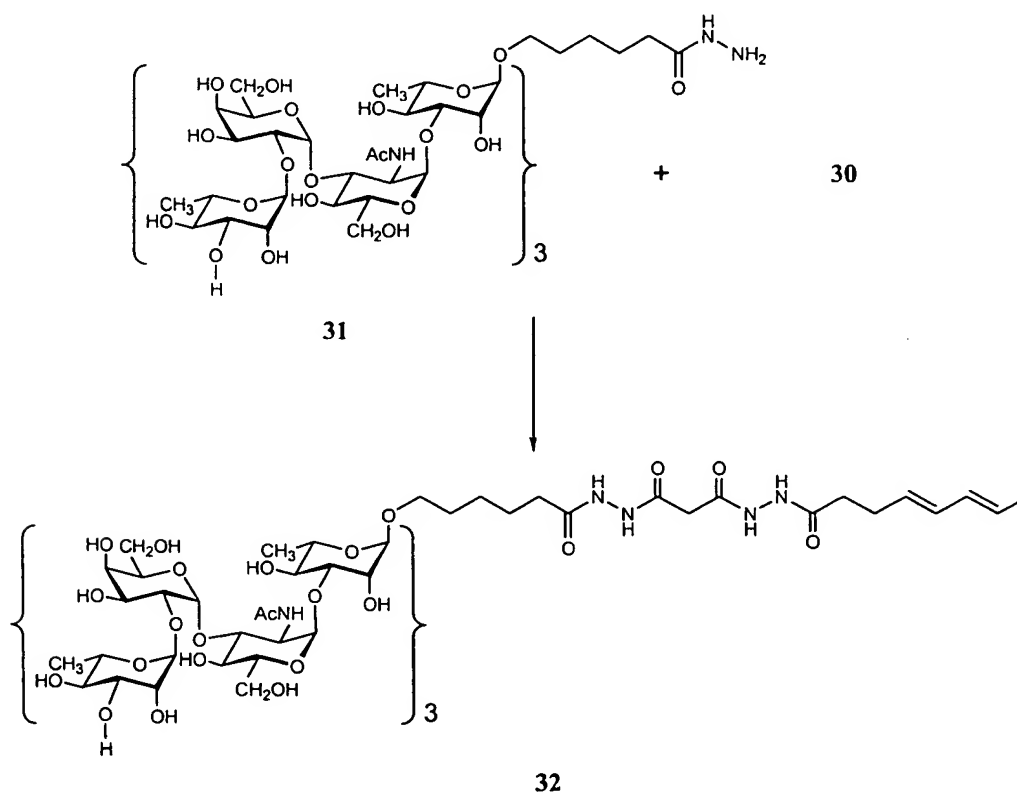


Diene component: The previously described ester 16 was converted to
 10 hydrazide 17 by treatment with hydrazine in MeOH at 23 °C for 24 hr. : ¹H NMR (500 MHz,

CD₃OD): δ = 6.00 (m, 2 H), 5.57 (m, 1 H), 5.49 (m, 1 H), 2.33 (m, 2 H), 2.21 (m, 2 H), 1.70 (d, 3 H).

Acylation with methyl malonyl chloride in pyridine at -20 °C for 20 min afforded the ester **29** in 72% yield, ¹H NMR (500 MHz, CD₃OD): δ = 6.03 (m, 2 H), 5.58 (m, 2 H), 3.74 (s, 3 H), 3.41 (m, 2 H), 2.31-2.43 (m, 4 H), 1.23 (d, 3 H).

Hydrolysis with LiOH in methanol (23 °C, 1 hr), followed by acidification with 1 N HCl, gave the crystalline acid **30** in 74% yield, ¹H NMR (500 MHz, CD₃OD): δ = 5.97 (m, 2 H), 5.52 (m, 2 H), 3.26 (m, 2 H), 3.24-3.37 (m, 4 H), 1.66 (d, 3 H).



10

Treatment of the dodecasaccharide hydrazide **31** (V. Pozsgay, *J. Org. Chem.* 1998, 63:5983) with the linker **30** in DMF in the presence of HATU led to the diene-equipped construct **32**, which was purified by gel-filtration through a Biogel P-4 column using water as the eluant. ¹H NMR (500 MHz, D₂O): δ = 6.11 (m, 2 H), 5.72 (m, 1 H), 5.61 (m, 1 H), 5.60 (br s, 3 H), 5.11 (br s, 2 H), 5.08 (br s, 1 H), 5.06 (br s, 2 H), 5.04 (d, 2 H), 5.00 (d, 1 H), 4.81

(br s, 1 H), 1.71 (d, 3 H); FAB-MS (dithiothreitol-dithioerythritol, positive ion) 2363.9 (M + Na), calcd.: 2362.9.

Dienophile component: A stirred solution of human serum albumin in pH 7.5 buffer (1 ml) was treated at 5 °C (ice-bath) with 3-sulfosuccinimidyl 4-maleimidobutyrate (**1**)
5 as described above, to provide maleimido-derivatized protein containing an average of 22 maleimido groups per molecule of protein.

Coupling reaction: Under conditions similar to those used for the conjugation of constructs **8** and **27**, cycloaddition of **32** onto the maleimido-derivatized protein took place at a slower rate. After 2 hours, the average incorporation was 1.5 dodecasaccharide chains per
10 HSA molecule, and after 8 hours, the incorporation reached an average of 3.